Giemsa Staining Kit

(Cat# K1426-30, -500; May-Grunwald; Store at RT)

I. Introduction:

The Giemsa Staining Kit (May-Grunwald) is intended for use in the visualization of cells present in hematopoietic tissues and certain microorganisms. This kit may be used on formalin-fixed, paraffin-embedded or frozen sections.

Nuclei: Blue/Violet; Cytoplasm: Light Blue; Collagen: Pale Pink; Muscle Fibers: Pale Pink; Erythrocytes: Gray, Yellow or Pink; Rickettsia: Reddish-Purple; Helicobacter Pylori: Blue; Mast Cells: Dark Blue with Red Granules

II. Application:

- Histological applications (IHC-Fr, IHC-P)
- For *in vitro* diagnostic use

III. Sample Type:

- Formalin-fixed, paraffin-embedded (5 microns) or frozen sections.
- Control Tissue: Blood Film, Bone Marrow, Spleen, or any well-fixed tissue.

IV. Kit Contents:

Components	K1426-30	K1426-500	Part Number	Storage Temperature
May-Grunwald Stock Solution	30 ml	500 ml	K1426-XX-1	RT
Giemsa Stock Solution	8 ml	500 ml	K1426-XX-2	RT
Phosphate Buffer Solution, pH 6.8	2 x 60 ml	500 ml	K1426-XX-3	RT

V. User Supplied Reagents and Equipment:

- Distilled water
- Coplin jars
- Xylene/Xylene Substitute
- Synthetic resin

VI. Shipment and Storage:

All the reagents are shipped and stored at room temperature (RT).

VII. Reagent Preparation:

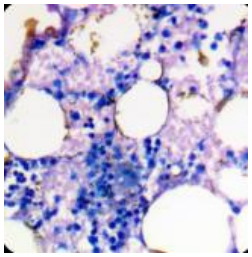
- Prepare Working May-Grunwald Solution by mixing 25 ml of May-Grunwald Solution with 25 ml of PBS Solution, pH 6.8
- Prepare Working Giemsa Solution by mixing 2.5 ml of Giemsa Stock Solution with 50 ml of PBS, pH 6.8
- Do not use past expiration date.
- Use caution when handling reagents.

VIII. <u>Procedure (Standard):</u>

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place slide in staining tray and flood with Working May-Grunwald Solution for 5-7 minutes. *Agitate slide occasionally to insure proper staining.*
3. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
4. Flood slide with Working Giemsa Solution for 10-15 minutes. *Note: Agitate slide occasionally to insure proper staining.*
5. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
6. Allow slide to remain in Phosphate Buffer Solution, pH 6.8 for an additional 3 minutes.
7. Dip slide quickly in distilled water to remove buffer and air dry at room temperature.
8. Dip slide twice in Xylene or Xylene Substitute.
9. Mount in synthetic resin.

<u>Procedure (Mast Cells):</u>

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place slide in staining tray and flood with Working May-Grunwald Solution for 5-7 minutes. *Note: Agitate slide occasionally to insure proper staining.*
3. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
4. Flood slide with Working Giemsa Solution for 10-15 minutes. Note: Agitate slide occasionally to insure proper staining.
5. Carefully flood slide with Phosphate Buffer Solution, pH 6.8 until stain no longer runs off.
6. Differentiate by dipping slide in Acetic Acid Solution (0.25%) until background is desired intensity.
7. Dip slide for 10 seconds in Phosphate Buffer Solution, pH 6.8 while agitating gently.
8. Dip slide quickly in distilled water to remove buffer and air dry at room temperature.
9. Dip slide twice in Xylene or Xylene Substitute.
10. Mount in synthetic resin.



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FOR RESEARCH USE ONLY! Not to be used on humans.