Sulfaquinoxaline ELISA Kit
(Catalog # E4773-100; 96 assays, Storage at 4°C)

I. Introduction:
Sulfaquinoxaline is a veterinary medicine that can be given to cattle and sheep to treat coccidiosis. It is an antiprotozoal agent used to combat coccidial infections of swine, cattle, fowl, and other veterinary animals. It is also used in controlling outbreaks of fowl typhoid and fowl cholera and in the treatment of infectious enteritis. BioVision Sulfaquinoxaline ELISA Kits based on Indirect-Competitive-ELISA principle. It can detect Sulfaquinoxaline in tissue, serum, honey, milk, urine samples. The micro-plate provided in this kit has been pre-coated with Sulfaquinoxaline. During the reaction, Sulfaquinoxaline in the samples or standard competes with Sulfaquinoxaline coated on the plate for binding to the anti-Sulfaquinoxaline antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microplate well, and TMB substrate is for color development. There is a negative correlation between the OD value of samples and the concentration of Sulfaquinoxaline. The concentration of Sulfaquinoxaline in the samples can be calculated by comparing the OD of the samples to the standard curve.

II. Applications:
In vitro, quantitative determination of Sulfaquinoxaline
Detection Range: Tissue (high detection limit) - 0.5 ppb, Tissue (low detection limit) - 2.5 ppb, Serum/urine - 2 ppb, Honey - 0.5 ppb, Milk - 10 ppb, Feed - 20 ppb
Sample recovery rate: Tissue/honey - 95% ± 25%, Urine/milk/serum/ feed - 85% ± 25%
Cross-reactivity: Sulfadiazine (SD or SDZ) <1%, Sulfamerazine (SM1) <1%, Sulfamethazine (SM2) <1%, Sulfamonomethoxine (SMM) <1%

III. Sample Type:
Tissue, Serum, Urine, Honey, Milk, Feed

IV. Kit Contents:

<table>
<thead>
<tr>
<th>Components</th>
<th>E4773-100</th>
<th>Part Number</th>
</tr>
</thead>
<tbody>
<tr>
<td>Micro ELISA Plate</td>
<td>96 wells</td>
<td>E4773-100-1</td>
</tr>
<tr>
<td>Standard</td>
<td>6 X 1 ml</td>
<td>E4773-100-2</td>
</tr>
<tr>
<td>HRP Conjugate</td>
<td>5.5 ml</td>
<td>E4773-100-3</td>
</tr>
<tr>
<td>Antibody Working Solution</td>
<td>5.5 ml</td>
<td>E4773-100-4</td>
</tr>
<tr>
<td>Substrate Reagent A</td>
<td>6 ml</td>
<td>E4773-100-5</td>
</tr>
<tr>
<td>Substrate Reagent B</td>
<td>6 ml</td>
<td>E4773-100-6</td>
</tr>
<tr>
<td>Stop Solution</td>
<td>6 ml</td>
<td>E4773-100-7</td>
</tr>
<tr>
<td>Wash Buffer (20X)</td>
<td>40 ml</td>
<td>E4773-100-8</td>
</tr>
<tr>
<td>Reconstitution Buffer (2X)</td>
<td>50 ml</td>
<td>E4773-100-9</td>
</tr>
<tr>
<td>Plate Sealer</td>
<td>3</td>
<td>E4773-100-10</td>
</tr>
</tbody>
</table>

V. User Supplied Reagents and Equipment:
- Microplate reader capable of measuring absorbance at 450 nm
- 0.1 M NaOH, HCl
- Clean Eppendorf tubes for preparing standards or sample dilutions

VI. Storage and Handling:
Store at 4°C.

VII. Reagent and Sample Preparation:
Bring all reagents to room temperature before use. Before using the kit, spin tubes and bring down all components to the bottom of tubes.

- **Wash Buffer (20X):** Dilute 20X Concentrated Wash Buffer to 1X with deionized water.
- **Reconstitution Buffer (2X):** Dilute 2X Reconstitution Buffer with deionized water. Mix 10x Reconstitution Buffer (V) : Deionized water (V) = 1:1. The Reconstitution buffer can be store at 4°C for a month.
- **0.2 M NaOH:** Dissolve 0.8 g NaOH with 100 ml deionized water
- **0.02 M PB Buffer:** Dissolve 2.58 g of Na2HPO4·12H2O and 0.44 g of NaH2PO4·2H2O to 500 mL of deionized water
- **0.5 M HCl:** Add 4.3 mL of concentrated HCl into 100 mL of deionized water and mix thoroughly
**VIII. Sample Preparation:**

**Sample pretreatment:**

- **Pretreatment of tissue (high detection limit):**
  Add 3±0.05 g of homogeneous tissue sample to a centrifuge tube, and then add 3 ml of 0.02 M PB buffer shake and mix thoroughly. Add 4 ml of ethyl acetate and 2 ml of acetonitrile, mix for 10 min, centrifuge at above 4000 rpm for 10 min. Take 2 ml of upper liquid (about 1 g of sample), blow-dry at 50-60 °C with nitrogen Evaporators/Water bath. Dissolve the residual with 1 ml of n-hexane, add 1 ml of Reconstitution buffer and shake for 1 min. Centrifuge at 4000 rpm for 5 min. Discard the upper n-hexane, take 50 μl of lower liquid for analysis.
  **Note:** Sample dilution factor: 1, minimum detection dose: 0.5 ppb

- **Pretreatment of tissue (low detection limit):**
  Add 2±0.05 g of homogeneous tissue sample to a centrifuge tube, then add 8 ml of 0.02 M PB buffer shake for 2 min, centrifuge at above 4000 rpm for 10 min. Take 50 μl of liquid for analysis.
  **Note:** Sample dilution factor: 5, minimum detection dose: 2 ppb

- **Pretreatment of serum:**
  Put blood sample at room temperature for 30 min, centrifuge at above 4000 rpm for 10 min, separate the serum or filter the serum. Take 1 ml of serum, add 3 ml of 0.02 M PB buffer, and mix for 30 sec. Take 50 μl of liquid for analysis.
  **Note:** Sample dilution factor: 4, minimum detection dose: 2 ppb

- **Pretreatment of honey:**
  Weigh 1±0.05 g of honey sample into 50 ml tube, add 8 ml of 0.5 M HCl solution, put at 37 °C for 30 min. Add 2.5 ml of 0.2 M NaOH solution (adjust the PH value to about 5), then add 4 ml of ethyl acetate, shake for 5 min, centrifuge at above 4000 rpm at room temperature for 10 min. Take 2 ml of upper liquid, blow-dry at 50-60 °C with nitrogen or air. Add 0.5 ml of Reconstitution buffer and mix for 30 sec. Take 50 μl of liquid for analysis.
  **Note:** Sample dilution factor: 1, minimum detection dose: 0.5 ppb

- **Pretreatment of urine:**
  Mix 3 ml of 0.02 M PB buffer solution and 1 ml of centrifuged clear urine sample for 30 sec. Take 50 μl of liquid to analysis.
  **Note:** Sample dilution factor: 4, minimum detection dose: 2 ppb

- **Pretreatment of milk:**
  Dilute milk sample with 0.02 M PB buffer solution with the ratio of 1:20 (for example, 20 μl milk + 380 μL of 0.02 M PB buffer), mix for 30 sec. Take 50 μl of liquid to analysis.
  **Note:** Sample dilution factor: 20, minimum detection dose: 10 ppb

- **Pretreatment of feed:**
  Weigh 2 g of crushed homogenate into the 50 ml tube, add 8 ml of acetonitrile, oscillate for 5 min, centrifuge at 4000 rpm for 5 min at room temperature. Take 1 ml of upper organic phase to another tube and blow-dry with Nitrogen Evaporators/Water bath at 50-60 °C. Add 1 ml of n-hexane, oscillate for 30 sec. Then add 1 ml of 0.2 M PB buffer solution, shake for 30 sec, and centrifuge at 4000 rpm for 5 min at room temperature. Remove the upper organic phase; take 100 μl of upper water phase to another tube. Add 900 μl of 0.2 M PB buffer solution; shake for 5 min to mix well. Take 50 μl for detection and analysis.
  **Note:** Sample dilution factor: 40, minimum detection dose: 20 ppb

**IX. Assay Protocol:**

**Note:** Bring all reagents and samples to room temperature 30 minutes prior to the assay. It is recommended that all standards and samples be run at least in duplicate. A standard curve must be run with each assay.

1. Add 50 μl of each **standard or samples** into appropriate wells.
2. Add 50 μl of **HRP Conjugate** and 50 μl of **Antibody Working Solution** to each well. Cover the plate with the sealer provided in the kit. Gently mix and incubate for 45 min. at 25°C.
3. Aspirate the solution from each well add 300 μl of **1x wash buffer** to each well. Leave it for 30 sec, aspirate the solution from each well and pat it dry against clean absorbent paper. Repeat this wash step 5 times.
4. Add 50 μl of **Substrate Reagent A** to each well and then add 50 μl of **Substrate Reagent B**. Cover with a new plate sealer. Incubate for about 15 min at 25°C. Protect the plate from light. **Note:** the reaction time can be shortened or extended according to the actual color change, but not more than 30 min.
5. Add 50 μl of **Stop Solution** to each well. Note: adding the stop solution should be done in the same order as the substrate solution.
6. Read the absorbance in micro plate reader set to 450 nm reference wavelength 630 nm. This step should be performed within 5 min after stop reaction.
X. Calculation:

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.

\[ \text{Absorbance (\%)} = \frac{A}{A_0} \times 100\% \]

A: Average absorbance of standard or sample  
\( A_0 \): Average absorbance of 0 ppb Standard

Typical standard curve and data is provided below for reference only. A standard curve must be run with each assay.

<table>
<thead>
<tr>
<th>Concentration of standard (ppb)</th>
<th>OD-1</th>
<th>OD-2</th>
<th>Average OD</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>2.0638</td>
<td>2.0463</td>
<td>2.0551</td>
</tr>
<tr>
<td>0.5</td>
<td>1.5246</td>
<td>1.5345</td>
<td>1.5296</td>
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<tr>
<td>1.5</td>
<td>1.1523</td>
<td>1.0860</td>
<td>1.1192</td>
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<tr>
<td>4.5</td>
<td>0.7814</td>
<td>0.7918</td>
<td>0.7866</td>
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<tr>
<td>13.5</td>
<td>0.4498</td>
<td>0.4585</td>
<td>0.4542</td>
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<tr>
<td>40.5</td>
<td>0.1665</td>
<td>0.1824</td>
<td>0.1745</td>
</tr>
</tbody>
</table>

XI. RELATED PRODUCTS:

- Diazepam ELISA Kit (E4772)
- Cimaterol ELISA Kit (E4771)
- Ciprofloxacin (Cipro) ELISA Kit (E4365)
- Enrofloxacin (ENR) ELISA Kit (E4277)
- Chloramphenicol (CAP) ELISA Kit (K4230)

*FOR RESEARCH USE ONLY! Not to be used on humans.*