Total Aflatoxin ELISA Kit
(Catalog # E4747-100; 96 assays; Storage at 4°C)

I. Introduction:
BioVision’s Total Aflatoxin ELISA Kit is based on the Competitive ELISA method. It can detect Aflatoxin (AF) in samples, such as grain, peanut, formula feed, etc. The microplate provided in this kit has been pre-coated with coupled antigen. During the reaction, AF in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-AF antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each micro plate well, and TMB substrate is added for color development. There is a negative correlation between the OD value of samples and the concentration of AF. The concentration of AF in the samples can be calculated by comparing the OD of the samples to the standard curve.

II. Applications:
• Quantitative measurement of Total Aflatoxin in grain and feed samples.
• Sample recovery rate: Grain/Formula feed - 85%±15%, Peanut- 82%±15%, Edible oil- 85%±15%, Sauce/Wheat/Barley feed-83%±15%, Beer- 84%±15%, Wine/Soy sauce/Vinegar- 87%±15%
• Detection limit: Grain- 0.1 ppb; Formula feed- 0.2 ppb; Edible oil/Peanut- 0.2 ppb; Sauce/Wheat/Barley feed- 0.2 ppb; beer- 0.2 ppb; Wine/Soy sauce/Vinegar- 0.1 ppb
• Sensitivity: 0.02 ppb (ng/ml)
• Cross-reactivity: Aflatoxin B1 (AFB1) - 100%, AflatoxinB2 (AFB2) - 80%, AflatoxinG1 (AFG1)- 75%, AflatoxinG2 (AFG2) - 45%, FlatoxinM1 (AFM1) - 8%

III. Sample Type:
• Wine, Soy, sauce, vinegar, Grain, Peanut, Formula feed, Edible oil

IV. Kit Contents:

<table>
<thead>
<tr>
<th>Components</th>
<th>E4747-100</th>
<th>Part Number</th>
</tr>
</thead>
<tbody>
<tr>
<td>ELISA Microplate</td>
<td>96 wells</td>
<td>E4747-100-1</td>
</tr>
<tr>
<td>Standards (S1 - S6)</td>
<td>1 ml X 6</td>
<td>E4747-100-2</td>
</tr>
<tr>
<td>HRP Conjugate</td>
<td>5.5 ml</td>
<td>E4747-100-3</td>
</tr>
<tr>
<td>Antibody Working Solution</td>
<td>5.5 ml</td>
<td>E4747-100-4</td>
</tr>
<tr>
<td>Substrate Reagent A</td>
<td>6 ml</td>
<td>E4747-100-5</td>
</tr>
<tr>
<td>Substrate Reagent B</td>
<td>6 ml</td>
<td>E4747-100-6</td>
</tr>
<tr>
<td>Stop Solution</td>
<td>6 ml</td>
<td>E4747-100-7</td>
</tr>
<tr>
<td>Wash Buffer (20x)</td>
<td>40 ml</td>
<td>E4747-100-8</td>
</tr>
<tr>
<td>Plate Sealer</td>
<td>3</td>
<td>E4747-100-9</td>
</tr>
</tbody>
</table>

V. User Supplied Reagents and Equipment:
• 70% Methanol.
• Deionized or distilled water
• Microplate reader with 450 nm wavelength filter

VI. Storage Conditions and Reagent Preparation:
Please store the opened kit at 4°C, protect from light and moisture and use within 2 months.

Wash Buffer: Dilute 20x Concentrated Wash Buffer with deionized water. Mix 1 ml of 20x Concentrated wash Buffer in 19 ml Deionized water to prepare 20 ml of 1X wash buffer (1:19).

Standard Concentration:

<table>
<thead>
<tr>
<th>Standards</th>
<th>S1</th>
<th>S2</th>
<th>S3</th>
<th>S4</th>
<th>S5</th>
<th>S6</th>
</tr>
</thead>
<tbody>
<tr>
<td>Concentration (ppb)</td>
<td>0</td>
<td>0.02</td>
<td>0.04</td>
<td>0.08</td>
<td>0.16</td>
<td>0.32</td>
</tr>
</tbody>
</table>

Sample Pretreatment:

Pretreatment of beer:
(1) Stir beer thoroughly to remove CO₂, take 2 mL of beer sample and add 1 mL of deionized water, then add 7 mL methanol, oscillate for 5 min;
(2) Take 0.5 mL of mixed sample liquid and add 0.5 mL of deionized water, mix well;
(3) Take 50 μL for detection and analysis. **Note: Sample dilution factor: 10, minimum detection dose: 0.2 ppb**
Pretreatment of Sauce/Wheat/Barley feed:
(1) Weigh 2 g of crushed homogenate into the 50 mL EP tube, add 10 mL of 70% methanol, oscillate for 5 min, centrifuge at 4000 rpm for 10 min at room temperature
(2) Take 2 mL of supernatant, add 4 mL of trichloromethane or dichloromethane, shake for 5 min, centrifuge at 4000 rpm for 10 min at room temperature
(3) Take the upper phase liquid to another tube; keep the lower phase liquid for use (lower liquid A). Add 4 mL of trichloromethane or dichloromethane to the upper liquid, shake well for 5 min, centrifuge at 4000 rpm for 10 min at room temperature. Discard the upper liquid and keep the lower liquid (lower liquid B)
(4) Mix lower liquid A and lower liquid B thoroughly
(5) Take 2 mL of mixed lower liquid and blow-dry with nitrogen evaporators/water bath at 50-60°C.
(6) Add 0.5 mL of 70% methanol to dissolve thoroughly, add 0.5 mL of deionized water, mix fully
(7) Take 50 μL for detection and analysis. Note: Sample dilution factor: 10, minimum detection dose: 0.2 ppb

Pretreatment of formula feed:
(1) Weigh 2 g of crushed homogenate into the 50 mL tube, add 10 mL of 70% methanol, shake for 5 min, centrifuge at 4000 rpm for 10 min at room temperature
(2) Take 0.5 mL of supernatant, add 0.5 mL of deionized water, mix well.
(3) Take 50 μL for analysis. Note: Sample dilution factor: 10, minimum detection dose: 0.2 ppb (If aflatoxin content is higher in the sample, take the mixed liquid from step 2, diluted with 35% methanol, the sample dilution multiple is the actual dilution multiple in that case. For example: take the mixed liquid from step 2, diluted 10 times with 35% of methanol, the actual dilution multiple is 10×10=100, detection limit: 2 ppb)

Pretreatment of grain:
(1) Weigh 2 g of crushed homogenate into the 50 mL tube, add 5 mL of 70% methanol, shake for 5 min, centrifuge at 4000 rpm for 10 min at room temperature
(2) Take 0.5 mL of supernatant, add 0.5 mL of deionized water, mix well.
(3) Take 50 μL for detection and analysis. Note: Sample dilution factor: 5, minimum detection dose: 0.1 ppb

Pretreatment of edible oil, peanut, high fat formula feed:
(1) Weigh 2 g of crushed homogenate into the 50 mL tube, add 8 mL of N-hexane and 10 mL 70% of methanol, oscillate for 5 min, centrifuge at 4000 rpm for 10 min at room temperature
(2) Discard the upper liquid, and take 0.5 mL of lower liquid, add 0.5 mL of deionized water, mix well.
(3) Take 50 μL for detection and analysis. Note: Sample dilution factor: 10, minimum detection dose: 0.2 ppb

Pretreatment of wine, soy sauce, vinegar:
(1) Take 2 mL of sample and add 2 mL of deionized water, then add 10 mL of trichloromethane or dichloromethane, shake for 5 min, centrifuge at 4000 rpm for 10 min at room temperature
(2) Take 1 mL of lower liquid and add 8 mL of N-hexane and 10 mL of 70% methanol, oscillate for 5 min, centrifuge at 4000 rpm for 10 min at room temperature
(3) Add 0.5 mL of 70% methanol to dissolve thoroughly, add 0.5 mL of deionized water, and mix well.
(4) Take 50 μL for detection and analysis. Note: Sample dilution factor: 5, minimum detection dose: 0.1 ppb

VII. Assay Protocol:
Restore all reagents and samples to room temperature before use. All the reagents should be mixed thoroughly by gentle mixing before pipetting. Avoid bubble formation.
It is recommended that all standards and samples be run at least in duplicate. A standard curve must be run with each assay.

1. Add 50 μL of standard or sample per well, then add 50 μL of antibody working solution, cover the plate sealer, shake for 5 sec gently to mix thoroughly, incubate in dark for 30 min at 37°C.
2. Remove the sealer carefully. Aspirate the liquid. Immediately add 300 μL of wash buffer (1x) to each well and wash. Repeat wash procedure for 5 times, 30 s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to remove them).
3. Add 100 μL of HRP conjugate to each well, incubate in dark for 30 min at 37°C.
4. Repeat wash step as described in step 2
5. Add 50 μL of substrate reagent A to each well, and then add 50 μL of substrate reagent B. Gently mix to mix thoroughly. Incubate at 37°C for 15 min in dark. Note: the reaction time can be shortened or extended according to the actual color change, but not more than 30 min.
6. Add 50 μL of stop solution to each well, shake the plate gently to mix thoroughly.
7. Determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. Read the plate within 5 min after adding stop solution.

8. Calculation: Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.
Absorbance (%) = \frac{A}{A_0} \times 100\%

A: Average absorbance of standard or samples
A_0: Average absorbance of 0 ppb Standard

VIII. Related Products:
- Aflatoxin M1 ELISA Kit (E4566)
- Aflatoxin B1 (AFB1) ELISA Kit (K4208)

FOR RESEARCH USE ONLY! Not to be used on humans.