Protein L Magnetic Beads

CATALOG #: 6537-1

AMOUNT: 1 ml

LOT #: ______________________

PREPARATION: Protein L Magnetic Beads are prepared by covalently coupling Recombinant Protein L (contains five Ig kappa light chain binding domains, BV catalog # 6530) to 6% crosslinked magnetically beaded agarose. The coupling technique is optimized to give a high binding capacity. The capacity of IgG binding is generally greater than 10 mg of human IgG per ml of wet gel.

CONTENTS: Supplied as a 50% slurry in 20 % Ethanol/dW.

TECHNICAL SPECIFICATIONS:

Parameter	Description
Support Characteristics	Paramagnetic, spherical, 6 % cross-linked agarose
Ligand	Recombinant Protein L
Particle Size	75 – 150 µm
Binding Capacity	Generally >10 mg human IgG/ml wet beads
Working Temperature	Room temperature
Storage Solution	20 % Ethanol/dW
Storage Temperature	4 – 8 °C
Stability	Stable, as supplied, for at least 1 year

FEATURES: Easy to use, high-binding capacity, non-adherent beads. Useful for immunoprecipitation and enrichment of antibodies. High affinity for kappa-light chain containing Ig antibodies from a variety of species. Protein L binds to all IgG subclasses from human, mouse and rat species. It also binds to human, mouse and rat IgM, IgA, IgE and IgD, as well as FAB with Kappa-light chains. Protein L is superior for binding chicken, hamster and pig IgG.

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USAGE: ***For Research Purpose Only! Not to be used in humans!***

Suggested Protocol:

Prepare the antibody solution by diluting the required amount of antibody in binding buffer before running the protocol.

1. Magnetic Bead Preparation (perform three times)
   a. Dispense the required amount of magnetic beads into a 1.5 ml microfuge tube.
   b. Place the tube in the magnetic rack and remove the storage solution.
   c. Add 500 µl binding buffer.
   d. Resuspend the beads.
   e. Remove the liquid
2. Antibody Capture
   a. Immediately add the antibody solution.
   b. Resuspend and mix (slow end-over-end) for at least 15 minutes.
   c. Remove the liquid.
3. Washing
   a. Add 500 µl Binding Buffer containing 0.5 M NaCl; Remove the liquid.
   b. Add 500 µl Binding Buffer; Remove the liquid.
4. Target Binding
   a. Add sample diluted in binding buffer.
   b. Incubate with slow end-over-end mixing for up to 60 minutes.
   c. Remove and collect unbound fraction.
5. Washing (perform three times)
   a. Add 500 µl wash buffer
   b. Remove liquid (save washes to troubleshoot)
6. Elution (perform three times)
   a. Add 2 volumes elution buffer (vs. bead volume).
   b. Completely resuspend beads and incubate at least 2 minutes.
   c. Remove and collect elution fraction.

Recommended Buffer Examples:

Binding buffer: 50 mM Tris, 150 mM NaCl, pH 7.5
Wash buffer: 50 mM Tris, 150 mM NaCl, pH 7.5 (or add 1% Octylglucoside to this buffer)
(Could also try 1X PBS as both binding and wash buffer)
Elution buffer: 0.1 M -0.2 M Glycine pH 2.5-3.1 (or 0.1 M citric acid, pH 2.5-3.1 or 2.5 % Acetic Acid)

RELATED PRODUCTS:

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Recombinant Protein G
Recombinant Protein L
Recombinant Protein A/G
Recombinant Protein A/G/L
Protein G Polyclonal Antibody
Protein A Polyclonal Antibody
Protein L Polyclonal Antibody
Protein A Sepharose
Protein G Sepharose
Protein L Sepharose
Protein A/G Sepharose
Protein A/G/L Sepharose
Protein G-Biotin
Protein G Coated Plates
Protein A Magnetic Beads
Protein G Magnetic Beads
Protein L Magnetic Beads
Protein A/G Magnetic Beads
Protein A/G/L Magnetic Beads
Protein G-FITC