

# Acid Phosphatase Assay Kit

(Catalog# K411-500; 500 reactions; Store kit at -20°C)

## I. Introduction:

Acid phosphatases (AP) dephosphorylate phosphate groups from phosphate esters under acid conditions. Different acid phosphatase isozymes are found in different organs, and their serum levels are used as a diagnostic for disease in the corresponding organs. Elevated prostatic acid phosphatase levels may indicate the presence of prostate cancer and elevated tartrate-resistant acid phosphatase levels may indicate the bone disease. BioVision's Acid Phosphatase Assay Kit is a high sensitivity, simple, direct and HTS-ready colorimetric assay designed to measure AP activity in serum and other samples. It is suitable for research and drug discovery. The kit uses *p*-nitrophenyl phosphate (*p*NPP) as a phosphatase substrate which turns yellow ( $\lambda_{max} = 405 \text{ nm}$ ) when dephosphorylated by AP. The kit can detect  $\mu\text{U}$  acid phosphatase activity in samples.

## II. Kit Contents:

Components	K411-500	Cap Code	Part No.
AP Assay Buffer	100 ml	NM	K411-500-1
<i>p</i> NPP Substrate (10 TAB)	1 vial	Red	K411-500-2
AP Enzyme	1 vial	Green	K411-500-3
Stop Solution	10 ml	WM	K411-500-4

## III. Storage and Handling:

Store the kit at -20°C, protect from light. Allow Assay Buffer to warm to room temperature before use. Briefly centrifuge vials before opening. Read the entire protocol before performing the assay.

## IV. Reagent Reconstitution and General Consideration:

***p*NPP Solution:** Dissolve 2 tablets *p*NPP into 5.4 ml Assay Buffer to generate 5 mM *p*NPP. Two tablets are sufficient for 100 assays. **NEVER TOUCH THE TABLETS WITH BARE HANDS.** The *p*NPP solution is stable for 12 hours on ice.

**AP Enzyme Solution:** Reconstitute AP Enzyme with 1 ml Assay Buffer. **NEVER FREEZE!** The enzymes are stable for up to 2 months at 4°C after reconstitution.

Ensure that the Assay Buffer is at room temperature before use. Keep samples, *p*NPP substrate solution, and AP Enzyme on ice during the assay.

## V. Acid Phosphatase Assay Protocol:

### 1. Sample Preparations:

Inhibitors of AP, such as tartrate, fluoride, EDTA, oxalate, and citrate, should be avoided in sample preparation. Serum, plasma, urine, semen, and cell culture media can be assayed directly. Cells ( $1 \times 10^5$ ) or tissue (~10 mg) can be homogenized in 100  $\mu\text{l}$  Assay Buffer, centrifuge to remove insoluble material at 13,000g, 3 minutes. Add test samples directly into 96-well plate, bring total volume to 80  $\mu\text{l}$  with Assay Buffer.

If samples contain color, it may interfere with O.D. 405 nm readings, use a sample background control. Add the same amount of sample into separate wells, bring volume to 80  $\mu\text{l}$ . Add 20  $\mu\text{l}$  stop solution and mix well to terminate AP activity in the sample.

2. Add 50  $\mu\text{l}$  of *p*NPP Substrate Solution to each well containing the test samples and background controls. Mix well. Incubate the reaction for 60 min at 25°C, protect from light.

### 3. Standard Curve:

Dilute 40  $\mu\text{l}$  of the 5 mM *p*NPP solution with 160  $\mu\text{l}$  Assay Buffer to generate 1 mM *p*NPP standard. Add 0, 4, 8, 12, 16, 20  $\mu\text{l}$  into 96-well plate in duplicate to generate 0, 4, 8, 12, 16, 20 nmol/well *p*NPP standard. Bring the final volume to 120  $\mu\text{l}$  with Assay Buffer.

Add 10  $\mu\text{l}$  of AP enzyme solution to each well containing the *p*NPP standard. Mix well. The AP enzyme will convert *p*NPP substrate to an equal amount of colored *p*-Nitrophenol (*p*NP). Incubate the reaction for 60 min at 25°C, protect from light.

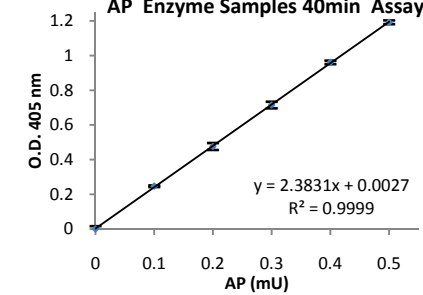
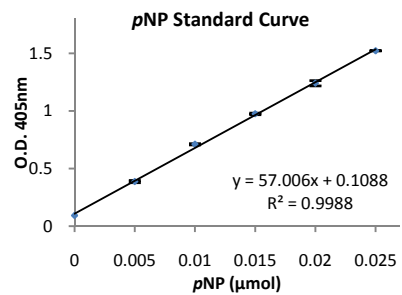
4. Stop all reactions by adding 20  $\mu\text{l}$  Stop Solution into each standard and sample reaction except the sample background control reaction (since 20  $\mu\text{l}$  Stop Solution has been added into the background control when the sample was prepared in step 1), gently shake the plate. Measure O.D. 405 nm in a micro-plate reader.

5. **Calculation:** Correct background by subtracting the value derived from the 0 standard from all the standards, samples and sample background controls (The background reading can be significant and must be subtracted from sample readings). Plot *p*NP standard Curve. Apply sample readings to the standard curve to get the amount of *p*NP generated by AP sample. AP activity of the test samples can then be calculated:

$$\text{AP activity (U/ml)} = A/V/T$$

Where A is amount of *p*NP generated by samples (in  $\mu\text{mol}$ ),  
 V is volume of sample added in the assay well (in ml),  
 T is reaction time (in minutes)

**Unit Definition:** One unit of AP is the amount of enzyme causing the hydrolysis of one micromole of *p*NPP to *p*NP per minute at pH 5.0 and 25°C.



## VI. Related Products:

- Alkaline Phosphatase Assay Kit
- Phosphate Fluorescence Assay Kit
- NAD/NADH Quantification Kit
- Lactate/Pyruvate Assay Kit/ II
- Ammonia Assay Kit
- Glucose Assay Kit

- ADP/ATP Ratio Assay Kit
- Phosphate Colorimetric Assay Kit
- Uric Acid Assay K
- Uric Acid Assay Kit
- Glutamate Assay Kit
- Fatty Acid Assay Kit